

Available online at www.sciencedirect.com



Tetrahedron 60 (2004) 3855–3862

Tetrahedron

# Monooxygenation of aromatic compounds by dioxygen with bioinspired systems using non-heme iron catalysts and tetrahydropterins: comparison with other reducing agents and interesting regioselectivity favouring meta-hydroxylation

Delphine Mathieu, Jean François Bartoli, Pierrette Battioni and Daniel Mansuy\*

UMR 8601, Université Paris V, 45 rue des Saints-Pères, 75270 Paris Cedex 06, France

Received 25 November 2003; revised 10 February 2004; accepted 2 March 2004

Abstract—Monooxygenation of aromatic compounds by dioxygen in the presence of catalytic amounts of an iron(II) salt and tetrahydropterins as reducing agents occurs with a regioselectivity favouring meta-hydroxylation of arenes bearing an electron-donating substituent, such as anisole, phenetole, toluene, and ethylbenzene. Comparison of similar systems using various reducing agents showed that only tetrahydropterins and ascorbate led to such a major meta-hydroxylation. The tetrahydropterin- and ascorbate-dependent systems should be useful for the preparation of meta-hydroxylated metabolites of aromatic drugs, as shown here in the case of diclofenac.  $© 2004 Elsevier Ltd. All rights reserved.$ 

#### 1. Introduction

Efficient systems based on iron porphyrins or non-heme iron complexes have been shown to mimic alkene epoxidation and alkane hydroxylation by cytochrome P450 and non-heme dependent monooxygenases.<sup>[1–5](#page-6-0)</sup> Cytochromes P450 and non-heme iron enzymes such as tetrahydrobiopterin (H4B)-dependent monooxygenases also catalyse the selective hydroxylation of aromatic compounds.<sup>1-6</sup> However, few model systems based on iron catalysts have been described so far for selective and efficient aromatic hydroxylation.<sup>[1–5](#page-6-0)</sup> The most efficient ones use  $H_2O_2$  as oxygen atom donor in the presence of either an iron porphyrin bearing electron-withdrawing  $\beta$ -substituents,<sup>7-9</sup> or a polyazamacrocyclic iron complex, Fe(tris-[N-(2 pyridylmethyl)-2-aminoethyl]amine)( $\text{ClO}_4$ )<sub>2</sub>, Fe(TPAA).<sup>[10](#page-6-0)</sup> Many systems using a Fe<sup>II</sup> salt,  $O_2$  as oxygen atom donor, and a reducing agent, such as ascorbate in the Udenfriend system, $^{11}$  $^{11}$  $^{11}$  have also been shown to be of interest for the hydroxylation of aromatic compounds (see for instance Ref.  $12-15$ ).

Tetrahydrobiopterin, H4B, is a necessary cofactor in non-heme iron aromatic aminoacid monooxygenases,<sup>[6](#page-6-0)</sup> and in NO synthases.[16](#page-6-0) In the latter enzymes, it plays a key role by transferring an electron to the heme inside the NOS active

0040–4020/\$ - see front matter © 2004 Elsevier Ltd. All rights reserved. doi:10.1016/j.tet.2004.03.006

site.[17](#page-6-0) These recent data led us to investigate the possible specific roles played by tetrahydropterins in monooxygenase model systems using non-heme iron complexes as catalysts,  $O_2$  as a source of oxygen atoms, and a reducing agent. A long time ago, preliminary experiments had shown that a system based on  $H_4B$  in the presence of catalytic amounts of  $Fe(SO<sub>4</sub>)<sub>2</sub>(NH<sub>4</sub>)<sub>2</sub>$  and EDTA was able to hydroxylate the aromatic ring of phenylalanine.<sup>[18](#page-6-0)</sup> Very recently, we have compared the properties of similar systems using different iron catalysts and reducing agents towards the hydroxylation of aromatic compounds. Here, we report that some of these systems using tetrahydropterins are efficient for aromatic hydroxylations, and show an unusual regioselectivity favouring the meta-hydroxylation of aromatic compounds bearing an electron-donating substituent.

#### 2. Results and discussion

## 2.1. Hydroxylation of anisole by a  $Fe^{II}/EDTA/O_2/$ tetrahydropterin system

Reaction of anisole with an aerobic phosphate buffer pH 7.4 containing H<sub>4</sub>B and catalytic amounts of Fe(SO<sub>4</sub>)<sub>2</sub>(NH<sub>4</sub>)<sub>2</sub> and EDTA, for 3 h at 20 $\degree$ C under conditions similar to those previously used by Viscontini et al. in the case of phenylalanine,<sup>[18](#page-6-0)</sup> led to a mixture of *ortho-*, *meta-* and para-methoxyphenols in a very unusual<sup>[9,10](#page-6-0)</sup> 36:46:18 ratio, and to minor amounts of phenol which should come from an

Keywords: Aromatic hydroxylation; Tetrahydrobiopterin; Iron salt; Diclofenac; Ascorbate.

<sup>\*</sup> Corresponding author. Tel.:  $+33142862187$ ; fax:  $+33142868387$ ; e-mail address: daniel.mansuy@univ-paris5.fr

<span id="page-1-0"></span>



Complete system: 1 mL aerobic 0.1 M phosphate buffer pH 7.4, containing Fe(SO<sub>4</sub>)<sub>2</sub>(NH<sub>4</sub>)<sub>2</sub>, EDTA and diMeH<sub>4</sub>P (molar ratio=1:8:16), and 1 mL anisole are vigorously stirred for 2 h at 20 °C; [iron catalyst]=1 mM. Sodium borohydride (10 equiv. relative to diMeH<sub>4</sub>P) was progressively added during the first hour to the reaction medium in order to regenerate diMeH<sub>4</sub>P, at least in part. o-OH, m-OH and p-OH are used for ortho-, meta- and para-methoxyphenol.



#### Scheme 1.

oxidative dealkylation of anisole. Identification of the products was performed by GC and confirmed by GC–MS. Similar results (Table 1) were obtained using the more accessible 6,7-dimethyltetrahydropterin, diMe $H_4P$ (Scheme 1), instead of H4B. However, replacement of  $H_4B$  with dihydrobiopterin,  $H_2B$ , led to an inactive system. In their  $H_4B$ -containing system, Viscontini et al.<sup>[18](#page-6-0)</sup> have described the beneficial effects on the yields of the addition of a co-reductant, sodium borohydride, which was supposed to regenerate H4B, at least in part, during the reaction. Table 1 shows that the addition of an excess of sodium borohydride to the  $Fe^{II}/EDTA/O$ <sub>2</sub>/diMeH<sub>4</sub>P system led to a marked increase of the aromatic hydroxylation yield (5.8 turnovers of the iron catalyst instead of 1.2). The absence of dioxygen (identical reaction under anaerobic conditions), or of the catalyst  $(Fe(SO<sub>4</sub>)<sub>2</sub>(NH<sub>4</sub>)<sub>2</sub>)$ , or of the reducing agents (diMeH<sub>4</sub>P+sodium borohydride), led to an almost inactive system (Table 1). Moreover, the absence of  $diMeH_4P$  alone led to a dramatic decrease of the yields, indicating that sodium borohydride is almost unable to act as an efficient reducing agent by itself and that its beneficial effects in the complete system is due to its regeneration of  $diMeH_4P$ during the reaction.

In Table 1 and the following tables, yields of hydroxylated products are generally expressed in turnovers of the iron catalyst (mol of product formed during the reaction per mol of catalyst). This parameter gives an idea of the catalytic efficiency of the system. From time to time, yields are also expressed in mol of product per mol of starting reducing agent (in %). This second parameter gives the number of electrons used by the system for dioxygen activation and hydroxylation of the aromatic substrates. It is an index of the decoupling between electron consumption by the system and hydroxylation of the substrate. In most bioinspired systems using an iron catalyst,  $O_2$  and a reducing agent, this decoupling is large and yields based on the reducing agent are low.<sup>[15](#page-6-0)</sup> In the system using both diMeH<sub>4</sub>P and sodium borohydride (in order to regenerate diMe $H_4P$ ), yields are

expressed relative to starting  $d$ iMeH<sub>4</sub>P, simply because sodium borohydride rapidly reacts with water and only a small, non measurable part of it is used to regenerate  $diMeH_4P.$ 

Kinetic experiments on the oxidation of anisole by the  $Fe^{II}/EDTA/O_2/diMeH_4P/NaBH_4$  system showed that the reaction was almost complete after 2 h, with an initial rate of methoxyphenols formation of 3.5 turnovers of the iron catalyst per hour (Fig. 1). Stopping of the reaction was presumably due to the consumption of the reducing agents, as a further addition of  $diMeH_4P$  and sodium borohydride led to similar yields and rates. Figure 1 shows that the regioselectivity of anisole hydroxylation varies as a function of the reaction time. At the beginning of the reaction  $(\sim 1 \text{ h})$ , the relative importance of meta-hydroxylation is at its maximum level, with an ortho:meta:para ratio of about 25:50:25, whereas this ratio tends towards 34:36:30 at the end of the reaction. This could mean that meta hydroxylation of anisole is favoured in the presence of the high concentrations of reductant that occur at the beginning of the reaction, and that could keep the Fe(II) concentration high.



Figure 1. Kinetic study of the formation of  $o$ -,  $m$ - and  $p$ -methoxyphenols upon oxidation of anisole by the  $Fe^{II}/EDTA/diMeH_4P/NaBH_4$  system (conditions of Table 1).





<sup>a</sup> Conditions as in the complete system described in [Table 1](#page-1-0).



Figure 2. Formula of the various chelates used in this study.

### 2.2. Studies of the reaction parameters

Table 2 shows the importance of the EDTA ligand on anisole hydroxylation by the above mentioned system. In the absence of EDTA, the system was almost unable to catalyze this reaction. Replacement of EDTA with analogous polydentate ligands, such as EGTA, EDDA, DTPA and  $HETA^{\dagger}$  (Fig. 2) led to a marked decrease of the methoxyphenols yields (0.2–3.2 turnovers/iron instead of 5.8 in the case of EDTA). This decrease of the yields was accompanied by changes of the regioselectivity which was still in favour of meta-methoxyphenol except in the case of DTPA. The distances between the coordinating groups of EDTA seem to play a crucial role, since replacement of EDTA with EGTA, which has a longer chain led to a dramatic decrease of the yields (Table 2).

A study of the effects of the Fe<sup>II</sup>/EDTA molar ratio on anisole hydroxylation showed that the best compromise for reaching good hydroxylation yields and a regioselectivity in favour of meta-hydroxylation was obtained with a 1:10 ratio (data not shown).

Replacement of  $Fe(SO<sub>4</sub>)<sub>2</sub>(NH<sub>4</sub>)<sub>2</sub>$  with the various non-heme iron complexes,  $[(TPAA)Fe^{II}](ClO<sub>4</sub>)<sub>2</sub>$ ,<sup>[10](#page-6-0)</sup>  $(L<sub>5</sub><sup>2</sup>FeCl)PF<sub>6</sub>$ <sup>[19](#page-6-0)</sup> and  $[Fe(TPA)(CH_3CN)_2]$ (ClO<sub>4</sub>)<sub>2</sub>,<sup>[20](#page-6-0)</sup> or with iron porphyrin complexes, such as Fe(III)[tetra-pentafluorophenyl- $\beta$ -tetra-



Moreover, a study of the effects of the concentration of the iron salt showed us that the yield of meta-methoxyphenol increased with the Fe<sup>II</sup> salt concentration up to  $0.5-1$  mM (Table 3). For concentrations higher than 1 mM, the yield of anisole hydroxylation dramatically decreased. Therefore, a Fe<sup>II</sup> concentration of 1 mM was used in the following studies.

Table 3. Effects of  $Fe<sup>H</sup>$  concentration on iron-catalyzed hydroxylation of anisole by  $O_2$  and diMeH<sub>4</sub> P<sup>a</sup>

$[Fe^{II}] (M)$	Aromatic hydroxylation						
	Total turnovers/catalyst	Regioselectivity $(\%)$					
		$o$ -OH	$m$ -OH	p-OH			
$4 \times 10^{-5}$ $5 \times 10^{-4}$ $10^{-3}$ $2 \times 10^{-3}$ $4 \times 10^{-3}$	0.8 6.9 5.8 0.7 0.2	80 39 30 29 50	10 30 43 43 25	10 31 27 28 25			

<sup>a</sup> Conditions as in the complete system described in [Table 1;](#page-1-0) molar ratio 1:8:16:160 for  $Fe^{II}/EDTA/diMeH_4P/s$  odium borohydride was conserved.

Finally, a study of the effects of pH on the hydroxylation of anisole showed that the total yield of methoxyphenols reach a maximum value at pH 7.4 (5.8 turnovers) ([Table 4\)](#page-3-0). However, the relative importance of meta-hydroxylation was enhanced when pH was increased from 4.5 up to 9, and was maximum at  $pH$  9 (52% of total methoxyphenols). The

 $\overline{\text{F}}$  EDTA, EGTA, EDDA, DTPA and HETA correspond to ethylenediamine tetraacetic acid, ethylenebis(oxyethylenenitrilo)-tetraacetic acid, ethylenediamine-N,N'-diacetic acid, diethylenetriamine pentaacetic acid and N(2-hydroxyethyl)ethylenediaminetriacetic acid respectively (see Fig. 2).

<span id="page-3-0"></span>**Table 4.** Effects of pH on iron(II)-catalyzed hydroxylation of anisole by  $O_2$ and diMe $H_4$   $P^a$ 

pH	Aromatic hydroxylation					
	Total turnover/catalyst	Regioselectivity (%)				
		$o$ -OH	$m$ -OH	p-OH		
4.5	1.1	54	31	15		
6	5.1	44	34	22		
7.4	5.8	30	43	27		
8.2	3.4	35	42	23		
9	2.5	29	52	19		
11.8	1.8	39	39	22		

<sup>a</sup> Conditions as in the complete system described in [Table 1,](#page-1-0) except for the buffer which was adapted to each pH (see Section 4).

following experiments were performed at pH 7.4 that was a good compromise for good yields and regioselectivity.

### 2.3. Generalization to various substrates

The Fe( $SO_4$ )<sub>2</sub>(NH<sub>4</sub>)<sub>2</sub>/EDTA/O<sub>2</sub>/diMeH<sub>4</sub>P system was also found to hydroxylate other aromatic compounds, such as benzene itself, with formation of the corresponding phenols (total turnovers of catalyst for aromatic hydroxylation between 1 and 5.8, and yields based on diMe $H_4P$  between 6.5 and 37%) (Table 5). The most distinctive and surprising characteristic of this system is the unusual regioselectivity of its hydroxylation of aromatic compounds bearing an electron-donating substituent. Thus, hydroxylation of anisole mainly occurred at the meta position (43% of total aromatic hydroxylation), and meta-hydroxylation of ethoxybenzene reached 38% of total aromatic hydroxylation. In contrast, the efficient iron catalyst/ $H_2O_2$  systems described previously $9,10$  almost exclusively led to *ortho*and para-hydroxylation of anisole and ethoxybenzene (*meta*-hydroxylation  $\leq 2\%$  of total aromatic hydroxylation), as expected for systems involving electrophilic hydroxylating species. With the present system, hydroxylation of the aromatic ring of toluene and ethylbenzene also occurred mainly at the meta position (50% of total aromatic hydroxylation, Table 5), whereas the previous iron catalyst/H<sub>2</sub>O<sub>2</sub> systems<sup>7–10</sup> gave much lower levels of *meta*hydroxylation for these two substrates (between 11 and 25%). This unusual regioselectivity of the diMe $H_4P$ dependent system was particularly well illustrated in the case of diclofenac (Scheme 2). This anti-inflammatory drug is metabolized by human cytochromes P450 with major formation of 4'-hydroxydiclofenac and minor formation of 5-hydroxydiclofenac, which are both derived from hydroxylations at the *para* positions of the strong electron-donating NH group.<sup>[23,24](#page-6-0)</sup> Iron porphyrin– $H_2O_2$  (or tBuOOH) systems mainly hydroxylate diclofenac at position 5, 4'-hydroxydiclofenac being formed in small amounts.[23](#page-6-0) In contrast, the diMeH4P-dependent system led to significant amounts of  $3'$ -hydroxydiclofenac, which results from meta-hydroxylation of the dichlorophenyl ring of diclofenac (32% of total aromatic hydroxylation, Table 5). This 3'-hydroxy metabolite, which was only formed in trace amounts with cytochromes P450<sup>[24](#page-6-0)</sup> and iron porphyrin model systems,<sup>[23](#page-6-0)</sup> was produced by the diMeH<sub>4</sub>Pdependent system in amounts comparable to those of 4'- and 5-hydroxydiclofenac (Table 5).

## 2.4. Effects of the replacement of  $\dim_{\mathbb{H}_{4}}P$  by other reducing agents

In order to better understand the origin of the important

**Table 5.** Regioselectivity of the hydroxylation of aromatic compounds by  $O_2$  and diMeH<sub>4</sub>P in the presence of Fe(SO<sub>4)2</sub>(NH<sub>4</sub>)<sub>2</sub> and EDTA<sup>a</sup>

Substrate	Aromatic hydroxylation				Other reactions yields (% based on diMeH <sub>4</sub> P) <sup>b,c</sup>	
		Total turnovers/catalyst Total yield (% based on $\text{d}\text{i}\text{MeH}_4\text{P}$ ) <sup>b</sup>	Regioselectivity $(\% )$			
			$o\text{-OH}$	$m$ -OH $p$ -OH		
Anisole	5.8	37	30	43	27	5 (phenol)
Ethoxybenzene	4	25	33	38	29	4 (phenol)
Toluene	2.8	17.5	40	50	10	$3$ (PhCH <sub>2</sub> OH)
Ethylbenzene		6.5	40	50	10	74 (PhCOCH <sub>3</sub> ) $+17$ (PhCHOHCH <sub>3</sub> )
Benzene		31				
Diclofenac	2.5	15.5		$32 (3'$ -OH): 36 (4'- OH: 32 (5-OH)		

Conditions as in the complete system described in [Table 1.](#page-1-0) In the case of diclofenac, saturating amounts of diclofenac sodium salt were dissolved in the phosphate buffer medium.

соон

Yields based on starting diMeH<sub>4</sub>P; one turnover of the catalyst roughly corresponds to 6% yield.<br>Reactions occurring in competition with aromatic hydroxylation are oxidative dealkylation for anisole and ethoxybenzene, an case of toluene and ethylbenzene.

 $O<sub>2</sub>$ , diMeH<sub>4</sub>P

Fe<sup>II</sup>, EDTA

3'-, 4'- and 5-hydroxydiclofenac in a 32/36/32 ratio

<span id="page-4-0"></span>**Table 6.** Effects of the nature of the reducing agent on iron-catalyzed hydroxylation of anisole by  $O_2^a$ 

Reducing agent	Aromatic hydroxylation							
	Total turnovers/catalyst	Total Yield <sup>b</sup> /reductant (% based)	Regioselectivity $(\%)$					
			$o$ -OH	$m$ -OH	$p$ -OH			
$DiMeH_4P+NaBH_4$	5.8	37(3.3)	30	43	27			
$DiMeH_4P$	1.2	7.5	35	43	22			
$H_4B$	0.9	5.5	36	46	18			
$H_4F$	1.5	9	39	33	28			
Ascorbate	6.4	40	31	40	29			
Catechol	0.7	4.5	44		56			
Hydroquinone	0.3	2	58		42			
Trimethylhydroquinone	2.9	17.5	39	22	39			
Thiophenol	< 0.2	$<$ 1						
$\alpha$ -Naphthol	< 0.2	<1						
Methylhydrazine	0.9	5.5	36	18	46			
2-Mercaptobenzoic acid	< 0.2	$<$ 1						
Hydrazobenzene	< 0.2	$\leq$ 1						
NADH	< 0.2	$<$ 1						

<sup>a</sup> Conditions as in the complete system described in [Table 1](#page-1-0), but without addition of sodium borohydride (except as specified for the first line of the table). Iron catalyst/reductant molar ratio=1/16

b Yields are based on the starting reducing agent; in the case of diMeH<sub>4</sub>P+Na borohydride, an excess of sodium borohydride was also used and yields are expressed on the basis of starting diMeH<sub>4</sub>P. Yield in parentheses is based on starting diMeH<sub>4</sub>P+sodium borohydride.

*meta*-hydroxylation observed with the Fe<sup>II</sup>/EDTA/O<sub>2</sub>/  $diMeH<sub>4</sub>P$  system, we have replaced diMe $H<sub>4</sub>P$  by various reductants. Some of them have been previously used in oxidizing systems based on an iron catalyst,  $O_2$  and a reducing agent. This is the case of catechol, $25$  hydro-quinone,<sup>[26](#page-7-0)</sup> trimethylhydroquinone,<sup>[27](#page-7-0)</sup> 2-mercaptobenzoic acid,<sup>[28](#page-7-0)</sup> hydrazobenzene<sup>[29](#page-7-0)</sup> and ascorbic acid.<sup>[11](#page-6-0)</sup> Table 6 shows that the regioselectivities the most in favour of metahydroxylation of anisole were obtained with tetrahydropterins diMe $H_4P$  and  $H_4B$ , and with ascorbate. Only these three reducing agents led to a major formation of metamethoxyphenol. Tetrahydrofolate,  $H_4F$  [\(Scheme 1](#page-1-0)), another natural tetrahydropterin cofactor, gave results similar to  $H_4B$  and diMeH<sub>4</sub>P, however with lower relative amounts of meta-methoxyphenol. Trimethylhydroquinone and methylhydrazine also led to significant amounts of meta-methoxyphenol but as a minor product when compared to its ortho and para isomers. Catechol and hydroquinone exclusively led to ortho- and para-methoxyphenol. All the other tested reducing agents, thiophenol,  $\alpha$ -naphthol, 2-mercaptobenzoic acid, hydrazobenzene and NADH failed to produce methoxyphenols in significant amounts. These data clearly showed that the structure of the reducing agent is a key factor in aromatic hydroxylations by the  $Fe^{II}/EDTA/O<sub>2</sub>/$ reductant systems. Two reducing agents emerge from this comparison, diMeH<sub>4</sub>P (or H<sub>4</sub>B) and ascorbate; they are much superior to the others both in terms of catalytic efficiency for anisole hydroxylation, with about six turnovers of the catalyst, and in terms of regioselectivity in favour of meta-hydroxylation (43 and 40% of total aromatic hydroxylation).

#### 2.5. Complementary mechanistic experiments

2.5.1. Origin of the oxygen atom inserted into substrates. Oxidation of anisole by  $^{18}O_2$  (containing 98%  $^{18}O$ ) and the  $Fe<sup>H</sup>/EDTA/diMeH<sub>4</sub>P$  system under conditions similar to those of [Table 1](#page-1-0) (see Section 4) led to ortho-, meta- and para-methoxyphenols bearing a phenolic oxygen atom containing  $98\pm1\%$  of <sup>18</sup>O isotope, as shown by mass

spectrometry coupled to gaz chromatography. When the same reaction was performed with  ${}^{16}O_2$  in H<sub>2</sub><sup>8</sup>O (98%) enriched), the methoxyphenol products contained less than  $1\%$  <sup>18</sup>O. These data clearly show that the oxygen atom incorporated into anisole almost exclusively came from  $O<sub>2</sub>$ (Scheme 3), whatever the position, *ortho*, *meta* or *para*, of the hydroxyl group.



Scheme 3.

**2.5.2. Possible role of**  $H_2O_2$ **.** All the iron/O<sub>2</sub>/reductant model systems of monooxygenases led to a partial decoupling between the consumption of electrons from the reducing agent and the monooxygenase reaction, which results in the formation of products coming from dioxygen reduction such as  $H_2O_2$ . In order to evaluate the role of H2O2 possibly produced during the studied reactions, oxidation of anisole by the  $Fe^{II}/EDTA/O_{2}/dMeH_{4}P$  system was performed in the presence of catalase. No significant difference was noted in the yields observed with or without catalase, implying that  $H_2O_2$  does not play an important role in these reactions.

#### 2.6. Discussion

Our data show that systems based on tetrahydropterins as reducing agents, in the presence of a catalytic amount of an iron(II) salt and EDTA, which are closely related to the system previously used by Viscontini et al. for the hydroxylation of phenylalanine, $18$  also catalyse the hydroxylation of many aromatic compounds such as benzene, anisole, ethoxybenzene, toluene, ethylbenzene and diclofenac. The main characteristic of these systems is their regioselectivity in favour of the meta-hydroxylation of

aromatic compounds bearing an electron-donating substituent. A comparative study of many systems using other reducing agents, under identical conditions, clearly showed that only  $diMeH_4P$  (or other tetrahydropterins) and ascorbate led to such a regioselectivity in favour of metahydroxylation. This suggests that a reinvestigation of metahydroxylation by Udenfriend-type systems  $(Fe^{II}/O<sub>2</sub>/$ ascorbate) should be very much interesting. In fact, there are few precedents of such iron-catalyzed meta-monooxygenations of aromatic compounds bearing an electrondonating substituent in the literature (for reviews, see Ref. [12–14](#page-6-0)). Most of the previously reported iron-catalyzed oxidations of such aromatic compounds, using either  $H_2O_2$ ,<sup>7-10,13,15</sup> or  $O_2$  and a reductant,<sup>12-15,30</sup> led to preferential ortho- and para-hydroxylations. Many reports on systems related to the one originally described by Udenfriend et al., and using  $O<sub>2</sub>$  and ascorbate, have been published; however very few mentioned meta-hydroxylation. Hamilton et al. have used a FeClO<sub>4</sub>/EDTA/O<sub>2</sub>/ ascorbate system and found at best 18% of metamethoxyphenol (relative to total aromatic hydroxylation) in the hydroxylation of anisole. $3<sup>1</sup>$  At the same period, Norman and Lindsay Smith<sup>13</sup> have used a similar system and reported regioselectivities for anisole and toluene involving 22 and 25% of meta-hydroxylated compound, respectively. More data have been published on metahydroxylation of aromatic compounds bearing an electrondonating substituent by another class of systems using Fe<sup>II</sup> or other reduced metal ions and dioxygen without any reducing agent.<sup>13,14,32-34</sup> These systems are not catalytic and yields are expressed relative to the metal ion. In the following, we will only consider the systems using a  $Fe<sup>II</sup>$  $salt^{13,32-34}$  for sake of comparison with our results exclusively based on  $Fe<sup>H</sup>$  catalysts. It has been noticed that the relative amounts of the meta-hydroxylated products increased when the iron concentration increased, and the regioselectivity the most in favour of meta-hydroxylation of anisole observed by Dearden et al. $34$  corresponded to a 39:50:11 o:m:p methoxyphenols ratio. These authors also mentioned that when they added N-benzyl-1,4-dihydronicotinamide as a reducing agent to their system, the meta-hydroxylated product almost completely disappeared.<sup>[34](#page-7-0)</sup>

The mechanisms of such *meta*-monooxygenations of aromatic compounds are presently almost unknown. The involvement of electrophilic species such as Fe<sup>III</sup>-OOH or high-valent iron–oxo intermediates, that are usually proposed in iron complex/ $H_2O_2$  systems, is unlikely, because these systems have been shown to mainly lead to ortho- and para-hydroxylated products.<sup>7-10,13,15</sup> Metahydroxylation does not seem to be due to the OH radical. because the regioselectivity observed for anisole hydroxylation by  $\cdot$ OH appears to correspond to a 84:0:16 molar ratio of the  $o$ -,  $m$ -,  $p$ -methoxyphenols.<sup>[13](#page-6-0)</sup>

The involvement of a polymetallic complex, containing at least two iron atoms, Fe–O and/or Fe–OO moieties, and the reducing agent, that would first attack the electronically favoured para (or ortho) position of the aromatic substrate and then its meta-position, would be at least partly in agreement with previous proposals from the literature.<sup>[13,34](#page-6-0)</sup> This would also fit with the increase of meta-hydroxylation upon increasing of the iron concentration, as previously

reported in some articles.<sup>[13,34](#page-6-0)</sup> In that regard, it is noteworthy that polymetallic iron complexes are formed upon reaction of  $Fe^{II}$  salts with EDTA,  $35$  and that some diiron complexes are able to catalyze the oxidation of toluene to meta-cresol, although in very different conditions.<sup>[36](#page-7-0)</sup> The reasons why only two of the tested reducing agents, diMeH<sub>4</sub>P (or H<sub>4</sub>B) and ascorbate, lead to a major meta-hydroxylation of anisole ([Table 6\)](#page-4-0), remain to be determined.

#### 3. Conclusion

Whatever its mechanism may be, the iron/EDTA/ tetrahydropterin/ $O<sub>2</sub>$  system is interesting because of its unusual regioselectivity favouring the incorporation of an oxygen atom from  $O_2$  at the *meta*-position of aromatic compounds bearing an electron-donating substituent. Our results show that this is true for many of such aromatic compounds. They also show that only two reducing agents, diMeH4P and ascorbate, used in the various  $Fe^{II}/EDTA/reduction/O<sub>2</sub>$  systems that we have compared, lead to this major meta-hydroxylation of anisole. Interestingly, the two corresponding systems also lead to the most efficient catalysis of anisole hydroxylation [\(Table 6](#page-4-0)).

Since a fast access to small amounts of various hydroxylated derivatives of drugs is required for identification of drug metabolites and for a first evaluation of the pharmacological properties of these metabolites, the aforementioned diMeH4P- and ascorbate-dependent systems should be useful for the preparation of meta-hydroxylated metabolites of aromatic drugs, as illustrated above in the case of diclofenac.

#### 4. Experimental

#### 4.1. Reactants

All the reactants and products were commercially available and purchased from Acros, Sigma, Aldrich and Alfa Aesar (iron salt). They were used without further purification except for the aromatic compounds that were eluted on a small alumina column prior to use.  $H_2^{18}O$  (98% enriched in <sup>18</sup>O) and <sup>18</sup>O<sub>2</sub> (98% enriched in <sup>18</sup>O) were purchased from EURISO-TOP (Saclay, France).

#### 4.2. General procedure

Hydroxylation of aromatic substrates was performed at room temperature in vials equipped with a magnetic stirrer. The organic phase consisting of 1 mL of aromatic substrate (anisole, phenetole, toluene, ethylbenzene or benzene) was added to 1 mL of 0.1 M phosphate buffer, pH 7.4, containing Fe(SO<sub>4</sub>)<sub>2</sub>(NH<sub>4</sub>)<sub>2</sub>·6H<sub>2</sub>O (10<sup>-3</sup> M, 10<sup>-6</sup> mol, 0.4 mg), EDTA  $(8\times10^{-3} \text{ M}, 8\times10^{-6} \text{ mol}, 2.3 \text{ mg})$ , diMeH<sub>4</sub>P  $(1.6 \times 10^{-2} \text{ M}, \quad 1.6 \times 10^{-5} \text{ mol}, \quad 3.8 \text{ mg})$  and NaBH<sub>4</sub>  $(1.6\times10^{-4} \text{ mol}, 6.4 \text{ mg})$ , the latter being progressively added in one hour. When other reductants than  $diMeH_4P$ were used,  $1.6\times10^{-5}$  mol was also introduced, but no sodium borohydride was added. After 2–3 h stirring at 20 °C, an internal standard (PhCOCH<sub>3</sub> or PhI,

<span id="page-6-0"></span> $1.6\times10^{-5}$  mol) was added and the reaction mixture was analyzed by gas chromatography.

A strictly similar procedure was followed when pH buffer was modified. In those cases, we used an acetate buffer adjusted to pH=4.5, a citric acid/Na<sub>2</sub>HPO<sub>4</sub> buffer pH=6, a trismabase/HCl buffer  $pH=8.2$ , a glycine/NaOH buffer  $pH=9$  and a Na<sub>2</sub>HPO<sub>4</sub>/NaOH buffer  $pH=11.8$ .

Reactions under anaerobic conditions were done by 'freeze–thaw cycles' of a vial containing all the reactants except the reducing agent, and of a second vial containing the reducing agent solution under argon. The content of the first vial was then transferred onto the reductant solution under argon.

#### 4.3. Hydroxylation of diclofenac

The aqueous phase (1 mL) contained  $Fe(SO<sub>4</sub>)<sub>2</sub>(NH<sub>4</sub>)<sub>2</sub>·6H<sub>2</sub>O$  $(10^{-3} M)$ , and EDTA  $(8 \times 10^{-3} M)$ , and 100 mg of the sodium salt of diclofenac were added to the solution. After 2–3 h stirring at 20 °C, samples were centrifuged to precipitate excess diclofenac, and products were analyzed by reverse phase HPLC, as described below.

## 4.4. Oxidation of anisole using  ${}^{18}O_2$

 $500 \mu L$  of  $0.1 M$  phosphate buffer, containing 7.0×10<sup>-7</sup> mol of Fe( $SO_4$ )<sub>2</sub>(NH<sub>4</sub>)<sub>2</sub>, 5.6×10<sup>-6</sup> mol of EDTA, and 1 mL of anisole were deaerated by three freeze–thaw cycles.  $1.1 \times 10^{-5}$  mol of diMeH<sub>4</sub>P were dissolved in 200  $\mu$ L of 0.1 M phosphate buffer (pH 7.4) and deaerated by the same procedure.  ${}^{18}O_2$  was introduced in the main vial and  $diMeH_4P$  was added using a deoxygenated syringe. After 2–3 h stirring, products were analysed by GC–MS as described below.

## 4.5. Oxidation of anisole using  $H_2^{18}O$

 $200 \mu L$  of 0.1 M phosphate buffer (pH 7.4) containing  $\text{Fe(SO}_4)_{2}(\text{NH}_4)_{2}$  (10<sup>-3</sup> M, 2×10<sup>-7</sup> mol) and EDTA  $(8\times10^{-3} \text{ M}, 1.6\times10^{-6} \text{ mol})$  were lyophilized and re-dissolved in 200  $\mu$ L of H<sub>2</sub><sup>8</sup>O. 500  $\mu$ L of anisole was added in the vial. diMeH<sub>4</sub>P (3.2 $\times$ 10<sup>-6</sup> mol) was introduced in the solid state, followed by  $N$ aBH<sub>4</sub>  $(1.6 \times 10^{-1}$  M,  $3.2 \times 10^{-5}$  mol), added in four times. After 2–3 h, products were analysed by  $GC-MS$  as in  ${}^{18}O_2$  experiments.

#### 4.6. Product analysis and identification

GC analyses were done using either a packed 5% FFAP (polar) column for anisole and benzene, or a capillary BP20 (polar) column for toluene and ethylbenzene, with detection with a flame ionization detector (FID). The products formed were analyzed by comparison of their retention time with those of authentic samples and by gas chromatography-mass spectrometry analysis using a Hewlett-Packard 5890 Series II GC coupled with a HP5972 mass selective detector.

HPLC analyses of diclofenac metabolites were done using a Chromatem 380 apparatus. Supernatant aliquots were injected onto a X-terra MS C18 column  $(3.0 \times 150 \text{ mm})$ ,  $5 \mu m$ ). The mobile phase (20 mM phosphate buffer (pH

8)/[acetonitrile/water (90/10)], gradient 20% up to 50% in 27 min) was delivered at a rate of 0.6 mL/min. Monitoring of the column effluent was performed with a detector at 270 and 280 nm. The reaction products were compared to authentic samples of  $3'$ -,  $4'$ - and 5-hydroxydiclofenac kindly provided by Ciba-Geigy (Basel, Switzerland).

#### References and notes

- 1. Meunier, B.; Robert, A.; Pratviel, G.; Bernadou, J. The porphyrin handbook; Kadish, K. M., Smith, K. M., Guilard, R., Eds.; Academic Press: New York, 1999; Vol. 4, p 119.
- 2. Mansuy, D. Coord. Chem. Rev. 1993, 125, 129.
- 3. Dolphin, D.; Traylor, T. G.; Xie, L. Acc. Chem. Res. 1997, 30, 251.
- 4. Costas, M.; Chen, K.; Que, L. Coord. Chem. Rev. 2000, 200– 202, 517.
- 5. Ménage, S.; Galey, J.-B.; Dumats, J.; Hussler, G.; Seité, M.; Gautier Luneau, I.; Chottard, G.; Fontecave, M. J. Am. Chem. Soc. 1998, 120, 13370.
- 6. Kappock, T. J.; Caradonna, J. P. Chem. Rev. 1996, 96, 2659.
- 7. Tsuchiya, N.; Seno, M. Chem. Lett. 1989, 263.
- 8. Carrier, M-N.; Sheer, C.; Gouvine, P.; Bartoli, J. F.; Battioni, P.; Mansuy, D. Tetrahedron Lett. 1990, 31, 6645.
- 9. Bartoli, J. F.; Le Barch-Ozette, K.; Palacio, M.; Battioni, P.; Mansuy, D. J. Chem. Soc., Chem. Commun. 2001, 1718.
- 10. Bartoli, J. F.; Lambert, F.; Morgenstern-Badarau, I.; Battioni, P.; Mansuy, D. C.R. Chimie 2002, 5, 263.
- 11. Udenfriend, S.; Clark, C. T.; Axelrod, J.; Brodie, B. B. J. Biol. Chem. 1954, 208, 731.
- 12. Hamilton, G. A. In Molecular mechanisms of oxygen activation; Hayaishi, O., Ed.; Academic: New York, 1974; p 405.
- 13. Norman, R. O. C.; Lindsay Smith, J. R. Oxidases and related redox systems; King, T. E., Mason, H. S., Morrison, M., Eds.; Wiley: Amherst, 1965; Vol. 1, p 131.
- 14. Ullrich, V.; Staudinger, Hj. Biological and chemical aspects of oxygenases: proceedings; Bloch, K. E., Hayaishi, O., Eds.; Maruzen Scientific: Tokyo, 1966; Vol. 1, p 235.
- 15. Metal-catalyzed oxidations of organic compounds; Sheldon, R. A., Kochi, J. K., Eds.; Academic Press: New York, 1981.
- 16. Pfeiffer, S.; Mayer, B.; Hemmens, B. Angew. Chem. Int. Ed. 1999, 38, 1714.
- 17. Hurschman, A. R.; Krebs, C.; Edmondson, D. E.; Huynh, B. H.; Marletta, M. A. Biochemistry 1999, 38, 15689.
- 18. Viscontini, M.; Mattern, G. Helvet. Chim. Acta 1970, 53, 372.
- 19. Bernal, I.; Jensen, I. M.; Jensen, K. B.; McKenzie, C. J.; Toftlund, H.; Tuchagues, J. P. J. Chem. Soc. Dalton Trans. 1995, 3667.
- 20. Zang, Y.; Kim, J.; Dong, Y.; Wilkinson, E. C.; Appelman, E. H.; Que, L., Jr. J. Am. Chem. Soc. 1997, 119, 4197.
- 21. Artaud, I.; Ben-Aziza, K.; Mansuy, D. J. Org. Chem. 1993, 58, 3373.
- 22. Stotter, D. A.; Thomas, R. D.; Wilson, M. T. Bioinorg. Chem. 1977, 7, 87.
- 23. Mancy, A.; Antignac, M.; Minoletti, C.; Dijols, S.; Mouries, V.; Duong, N. T.; Battioni, P.; Dansette, P. M.; Mansuy, D. Biochemistry 1999, 38, 14264.
- 24. Bort, R.; Macé, K.; Boobis, A.; Gomez-Lechon, M.-J.; Pfeifer, A.; Castell, J. Biochem. Pharmacol. 1999, 58, 787.

<span id="page-7-0"></span>

- 25. Hamilton, G. A.; Friedman, J. P. J. Am. Chem. Soc. 1963, 85, 1008.
- 26. Funabiki, T.; Yokomizo, T.; Suzuki, S.; Yoshida, S. Chem. Commun. 1997, 151.
- 27. Raffard, N.; Balland, V.; Simaan, J.; Létard, S.; Nierlich, M.; Miki, K.; Banse, F.; Anxolabéhère-Mallart, E.; Girerd, J.-J. C.R. Chimie 2002, 5, 99.
- 28. Ullrich, V. Z. Naturforschg 1969, 24b, 699.
- 29. Mimoun, H.; Seree de Roch, I. Tetrahedron 1975, 31, 777.
- 30. (a) Funabiki, T.; Toyoda, T.; Ishida, H.; Tsujimoto, M.; Ozawa, S.; Yoshida, S. J. Mol. Catal. 1990, 61, 235. (b) Kitajima, N.; Ito, M.; Fukui, H.; Moro-oka, Y. J. Chem. Soc., Chem. Commun. 1991, 102.
- 31. Hamilton, G. J. Am. Chem. Soc. 1964, 86, 3390.
- 32. Nofre, C.; Cier, A.; Lefier, A. Bull. Soc. Chim. France 1961, 530.
- 33. Staudinger, Hj.; Ullrich, V. Z. Naturforshg 1964, 19b, 877.
- 34. Dearden, M. B.; Jefcoate, C. R. E.; Lindsay Smith, J. R. Adv. Chem. Ser. 1968, 77, 260.
- 35. Mizuta, T.; Wang, J.; Miyoshi, K. Inorgan. Chim. Acta 1995, 230, 119.
- 36. Mukerjee, S.; Stassinopoulos, A.; Caradonna, J. P. J. Am. Chem. Soc. 1997, 119, 8097.